

Effect of radiation process followed by traditional treatments on nutritional and antinutritional attributes of sorghum cultivar

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Abstract

The present work investigates the effect of radiation on nutritional and antinutritional attributes of raw and processed flour of sorghum cultivar (WadAhmed). The flour of the cultivar was radiated using gamma radiation at doses of 5, 10 and 15 kGy and thereafter the radiated flour was fermented and/or cooked. Radiation of raw flour decreased the level of antinutrients with a concomitant increase in protein digestibility and minerals extractability. Moreover, radiation increased the level of some amino acids and decreased others. Among amino acids the most limiting ones are lysine, methionine plus cysteine and threonine. Fermentation and/or cooking of radiated and non-radiated flour reduced antinutrients and improved the protein digestibility, minerals extractability and level of some amino acids. Most of the amino acids were slightly stable against all treatments with few exceptions.

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Introduction

African people like to consume sorghum as fermented flat bread (Kisra), thick porridge (Aceda), thin fermented gruel (Nasha), boiled grains (Balella) and beverages (Abreh and Hulumur) as a source of protein and minerals (Abdelseed et al., 2011). Irradiation could replace or supplement chemical preservatives; in other cases it may have unique advantages whether dry or frozen foods (Sendra et al., 1996). In agricultural science and food technology, recent research has elucidated new potential applications for radiation. High doses of ionising radiation have been shown to inhibit growth of microbs (Mbarki et al., 2008; Bhavsar et al., 2007) and to reduce the level of antinutrients (Mohamed et al., 2010a) which were reported to reduce the availability of proteins and minerals (Idris et al., 2007; Osman et al., 2010; Mohiedeen et al., 2010; Mohamed Nour et al., 2010; Sokrab et al., 2012).

There are also many reports supporting the use of gamma irradiation as a fungicidal agent (Aziz *et al.*, 2007). Seeds of different plants that are consumed as food have varying nutrient values which are dependent on the basic constituents of seed proteins. The amino group $(-NH_2)$ is the most radiosensitive portion of the amino acids (Siddhuraju *et al.*, 2002). Extensive research showed that the macronutrients (carbohydrates, proteins and lipids) content are

relatively stable against irradiation dose sup to10 kGy (WHO, 1999). However, Lee *et al.* (2005) reported that gamma irradiation affects proteins by causing conformational changes, oxidation of amino acids, rupturing of covalent bonds and formation of protein free radicals. Also, chemical changes in the proteins caused by gamma irradiation include fragmentation, cross-linking, aggregation and oxidation by oxygen radicals that are generated in the radiolysis of water.

Gamma irradiation has a slight effect on the amino acid profile at recommended doses to foods (WHO, 1999). This effect could be related to the structure of each amino acid, simple amino acids increased upon irradiation, such as glycine, which undergo reductive deamination and decarboxylation (Erkanand and Ozden, 2007). In addition, aliphatic amino acids with increasing chain length provide additional C-H bonds for interaction with OH radicals which reduces the extent of oxidative deamination (Erkanand and Ozden, 2007). Wang and vonSonntage (1991) reported that sulphur-containing as well as aromatic amino acids are, in general, the most sensitive to irradiation, while simple amino acids could be formed by destruction of other amino acids. Mohamed et al. (2010a) reported that millet flour had a severe problem during storage and it was observed to produces off-flavour and bitter taste which can be overcome by using radiation.

The literature has many reports demonstrating that thermal processing methods improve the nutritional

quality of foods. However, there is a scarcity of information relating to the effects of processing with ionizing energy. Therefore, the present work was carried out to investigate the effect of radiation process followed by traditional treatments on nutritional quality of raw and processed sorghum cultivar flour.

Materials and Methods

Materials

Grains of the sorghum cultivar (WadAhmed) were collected from Department of Agronomy, Faculty of Agriculture, University of Khartoum, Sudan. Collected seeds (4 kg) of the cultivar were ground to pass a 0.4 mm screen. All chemicals used for the experiments were of reagent grade.

Samples preparation

The flour with a moisture content of 5.45% were spread uniformly and stored in polythene bags of mass of 100 gm, Gamma radiation process was conducted at Kaila irradiation processing unit, Sudanese Atomic Energy Corporation (SAEC). The flour was exposed to gamma rays generated by a cobalt-60 source (Gammacell 220, MDS Nordion, Ottawa, Canada). The flour was radiated at 0, 5, 10 and 15 kGy following the procedures described by Helinski et al. (2008) with a dose rate of ca. 3.2 kGy/h at $24 \pm 1^{\circ}$ C and normal relative humidity. Double side irradiation (exposure to both sides) was performed for uniform dose delivery. A dosimetry system was used to measure the dose received by the batch based on the Gafchromic HD-810 film (International Specialty Products, NJ, USA; FAO/IAEA/USDA 2003). Three dosimeters were included with each batch of flour and read after irradiation with a Radiachromics reader (Far West Technology Inc., CA, USA). All experiments were repeated 3 times and 3 replicates of each flour type were irradiated. Radiated and nonradiated flour of the cultivar was naturally fermented till the pH of the dough reached 4.50 and thereafter cooked for 20 min in a water bath and then dried and ground to pass a 0.4 mm screen for further analysis.

Tannin determination

Quantitative determination of tannins was carried out using the modified vanillin-HCl method according to Price and Butler (1978) using 200 mg sample. A standard curve was prepared expressing the results as catechin equivalents, i.e amount of catechin (mg per ml) which gives a colour intensity equivalent to that given by tannins after correcting for blank.

Phytic acid determination

Phytic acid content was determined by the

method described by Wheeler and Ferrel (1971) using 2.0 g dried sample. A standard curve was prepared expressing the results as $Fe(NO_3)_3$ equivalent. Phytate phosphorus was calculated from the standard curve assuming a 4:6 iron to phosphorus molar ratio.

In vitro protein digestibility determination

In vitro protein digestibility was carried out using single enzyme (pepsin) digestion according to the method of Maliwal (1983) in the manner described by Monjula and John (1991). Digestibility was calculated using the following equation:

Protein digestibility (%) = $\frac{N \text{ in supernatant} - N \text{ in pepsin } x 100}{N \text{ in sample}}$

Total and extractable minerals determination

For total content, minerals were extracted from the samples by the dry ashing method described by Chapman and Pratt (1982). Minerals were determined using atomic absorption spectrophotometer (Perkin-Elmer 2380). Na and K contents were determined using flame photometer (corning 400). For extractability, minerals in the samples were extracted by the method described by Chauhan and Mahjan (1988) using 1.0 g of the sample. HCl extractability (%) was determined as follows:

Amino acids composition determination

The amino acids composition of the samples was measured on hydrolysates using amino acids analyzer (Sykam-S7130, Tokyo, Japan) based on high performance liquid chromatography technique. Sample hydrolysates were prepared following the method of Moore and Stain (1963). About 200 mg of the sample was taken in a hydrolysis tube. Then 5 ml of 6 N HCl was added to the sample and the tube tightly closed and incubated at 110°C for 24 h. After incubation, the solution was filtered and 200 ml of the filtrate was evaporated to dryness at 140°C for 1 h. The hydrolysates after dryness were diluted with 1.0 ml of 0.12 N citrate buffer (pH 2.2). Aliquot of 150 µl of the sample hydrolysates was injected in an action separation column at 130°C.

Ninhydrin solution and an eluent buffer (solvent A, pH 3.45 and solvent B, pH 10.85) were delivered simultaneously into a high temperature reactor coil (16 m length) at a flow rate of 0.7 ml/min. The buffer/ninhydrin mixture was heated in the reactor at 130°C for 2 min to accelerate chemical reaction of amino acids with ninhydrin. The products of the reaction mixture were detected at wavelengths of 570 and 440 nm on a dual channel photometer. The amino

acids composition was calculated from the areas of standards obtained from the integrator and expressed as gm/100 gm protein.

Essential amino acid (EAA) score determination

The essential amino acid (EAA) score was determined by applying the formula:

AAS score % =
$$gm \text{ of EAA in 100 gm test protein}$$
 x 100
gm of EAA in 100 gm FAO/WHO reference pattern

Statistical analysis

Each determination was carried out on three separate samples and analysed in duplicate on dry weight basis; the figures were then averaged. Data were assessed by the analysis of variance (Snedecor and Cochran, 1987). Comparisons of means for treatments were made using Duncan's multiple range tests. Significance was accepted at $P \ge 0.05$.

Results and Discussion

Effect of gamma radiation and/or traditional processing on antinutrients and in vitro protein digestibility

The effect of gamma irradiation and/or traditional processings on tannin and phytate contents and in vitro protein digestibility of sorghum cultivar (WadAhmed) are shown in Table 1. Tannin content of raw flour significantly ($P \le 0.05$) reduced after radiation and the level of reduction increased with increase in radiation dose as reported by Mohamed et al. (2010a) for millet cultivars. Traditional processing (fermentation and/or cooking) of raw flour significantly ($P \le 0.05$) decreased the level of tannin which agree with that reported by Idris et al. (2005), who stated that combination of fermentation and cooking of sorghum improved the nutritional quality and drastically reduced tannin content to safe levels than any other processing methods. Also Osman (2004) reported that fermentation markedly reduced tannin content of sorghum cultivars. The reduction in tannin content due to fermentation is likely to be due to biochemical activity of fermenting organisms (Eltayeb et al., 2007). However, when traditional treatments were applied in combination with radiation process further reduction in tannin content was observed with a minimum value of 0.299% obtained for fermented and cooked seeds that radiated at a dose of 10 KGy. The results are in agreement with those reported by Brigide and Canniatti-Brazaca (2006) who observed that tannin content was inversely correlated with the applied irradiation doses. Radiation of raw flour significantly $(P \le 0.05)$ reduced the level of phytate and the rate of

Table 1. Tannin (%), Phytate (mg/100 g) and *in vitro* protein digestibility (IVPD) of sorghum cultivar Wad Ahmed as affected by irradiation followed by processing

Radiation dose	Treatment	Parameter							
(KGy)		Tannin	Phytate	IVPD					
0	Raw	1.400±0.190ª	268.00±1.50ª	51.6±0.12°					
	Fermented	0.930±0.205°	130.20±0.12 ^m	57.84±0.03 ^d					
	Cooked	0.615±0.014 ⁱ	211.72±0.18°	38.24±1.012 p					
	Fermented/cooked	0.379±0.035m	166.96±0.05 ⁱ	43.86±2.431					
5	Raw	1.282±0.051b	236.11±1.92b	53.72±0.08g					
	Fermented	0.866±0.023 ^f	113.58±0.12 ⁿ	59.50±0.07°					
	Cooked	0.559 ± 0.019^{j}	185.89±3.16 ^f	39.23±1.64°					
	Fermented/cooked	0.339±0.008 ⁿ	147.34±0.99 j	44.10±0.05k					
10	Raw	1.109±0.052°	220.76±0.11°	54.85±0.04 ^f					
	Fermented	0.754±0.153g	110.19±0.02°	60.63±0.10ª					
	Cooked	0.492±0.028k	179.54±0.10g	40.40±1.43 ⁿ					
	Fermented cooked	0.299±0.025°	141.37±2.01 ^k	44.53±0.08 ⁱ					
15	Raw	1.106 ± 0.115^{d}	220.28±0.06 ^d	55.49±0.21e					
	Fermented	0.722 ± 0.071 h	105.76±0.12 ^p	60.89±2.05 ^a					
	Cooked	0.464 ± 0.017^{1}	171.71 ± 0.10^{h}	40.70±0.01m					
	Fermented cooked	0.284±0.053p	135.24±0.291	44.69±0.09i					

superscript(s) in a column are significantly different at $p \le 0.05$.

reduction increased with increase in radiation dose as reported by Mohamed et al. (2010a) for millet cultivars. Traditional processing (fermentation and/ or cooking) significantly ($P \le 0.05$) decreased the level of phytate as reported by Idris et al. (2005), who found that combination of cooking and fermentation of sorghum significantly ($P \le 0.05$) reduced the level of phytate. Also Osman (2004) reported that fermentation markedly and significantly ($P \le 0.05$) reduced phytate content of sorghum cultivars. The reduction in phytate content due to fermentation is likely due to biochemical activity of the microbial enzyme phytase which hydrolyzed phytate. However, when traditional treatments applied in combination with radiation process further reduction was observed with a minimum value of 105.76 mg/100 gm obtained for fermented flour radiated at 15 KGy. Osman et al. (2012) reported that phytic acid and tannin contents of broad bean were significantly ($P \le 0.05$) reduced as a result of radiation. El- Niely (2007) stated that radiation after processing significantly ($P \le 0.05$) decreased the level of phytic acid of legumes. The reduction in phytic acid during radiation process is likely to be due to chemical degradation of phytate to the lower inositol phosphates and inositol by the action of free radicals produced by radiation or might be due to cleavage of the phytate ring itself (Siddhuraju et al., 2002). Both cereal and microbial phytases can contribute to a reduction in phytate during fermentation process (Eltayeb et al., 2007). Duodu et al. (1999) reported that cooking did not decrease phytic acid in sorghum porridge, but cooking and irradiation caused a significant decrease (40%). The effect of gamma irradiation on in vitro protein digestibility (IVPD) of WadAhmed cultivar is shown in Table 1. Radiation of raw flour was found to be effective in improving the IVPD at low dose of 5 KGy but as the level of radiation increased the IVPD significantly ($P \le 0.05$) decreased. Traditional processing (fermentation and/or cooking) of raw

Radiation dose (KGy)		Minerals										
	Treatment	N	la	K	-	Mg						
		Total	extractable	Total	extractable	Total	extractable					
0	Raw	18.10±1.95 ^{ef}	65.70±0.05 ⁿ	380.0 ± 0.01^{f}	51.60±0.09i	54.30±1.05 ⁱ	47.50±0.95°					
	Fermented	21.50±2.73b	78.90±1.95g	410.20 ± 1.94^{d}	69.11±0.61 ^d	58.00±1.19°	72.30±1.65g					
	Cooked	18.30±0.09e	66.90 ± 0.05^{k}	346.10±0.75 ⁿ	52.62 ± 0.01^{h}	54.00±1.59 ^j	45.10±0.02 ^p					
	Fermented/cooked	17.96±0.96 ^{ef}	77.00 ± 0.01^{i}	387.80±0.87e	72.65±0.42bc	55.80 ± 0.35^{f}	67.40±0.41 ^h					
5	Raw	18.10±1.89 ^{ef}	66.50 ± 1.32^{km}	377.50 ± 0.54^{g}	50.03±1.61 ^j	54.40 ± 0.08^{hi}	50.00±0.83 ⁿ					
	Fermented	21.00 ± 0.97^{b}	81.60±0.65 ^e	413.10±0.76°	68.91 ± 0.24^{d}	59.60±0.04ª	80.00±0.97e					
	Cooked	17.43 ± 0.23^{fg}	62.50±0.16°	$370.40{\pm}0.18^{i}$	51.93 ± 0.86^{hi}	52.00 ± 0.02^{k}	48.90±0.43 ⁿ					
	Fermented/cooked	20.27±0.37°	$78.20{\pm}0.85^{h}$	359.00 ± 0.76^{m}	73.53±0.45 ^b	56.30±0.01°	74.70±0.87f					
10	Raw	19.00±3.03 ^d	72.60±1.21 ^j	$380.00{\pm}0.68^{f}$	51.15±1.92 ⁱ	54.80 ± 0.01^{h}	56.00±0.52 ^j					
	Fermented	22.20±0.76ª	88.90±0.65ª	420.30±0.97 ^b	79.08±0.98ª	59.30±0.60 ^b	99.10±0.98ª					
	Cooked	17.78±0.43 ^f	79.90 ± 0.89^{f}	361.00 ± 0.76^{1}	53.69±0.54g	52.00±0.19k	52.50±0.23k					
	Fermented/cooked	21.32±0.53b	84.10±0.09°	365.00±0.34 ^j	78.74±0.23ª	57.20±0.12 ^d	81.90±0.96d					
15	Raw	18.31±2.05e	66.20±0.37 ^m	376.30 ± 0.54^{h}	55.28 ± 0.04^{f}	54.00±0.91 ^j	57.50±0.82 ⁱ					
	Fermented	20.17±0.43°	87.60 ± 0.87^{b}	424.30±0.17 ^a	78.19±0.75 ^a	59.70±0.74ª	95.00±0.60 ^t					
	Cooked	17.60 ± 0.67^{fg}	83.30 ± 0.65^{d}	363.00 ± 0.54^{k}	57.23±0.34e	54.00 ± 0.53^{j}	50.90±1.80 ¹					
	Fermented/cooked	18.20±0.29 ^{ef}	81.50±0.78 ^e	380.00 ± 0.87^{f}	72.16±0.12°	55.80±0.86 ^g	86.80±0.54					

Table 2. Total (mg/100 gm) and extractable (%) Sodium (Na), Potassium (K) and Magnesium (Mg) of Wad Ahmed cultivar as affected by gamma irradiation followed by processing

flour significantly (P ≤ 0.05) increased the level of IVPD as reported by AbdElhaleem et al. (2008). When traditional treatments applied in combination with radiation process further improvement in IVPD was observed with a maximum value of 60.89% for fermented flour radiated at 15 KGy. The results obtained agrees with results obtained by El-Niely (2007) who studied the influence of irradiation on in vitro protein digestibility of broad beans irradiated at 2.5, 5, 10 and 20 KGy, he observed that the in vitro protein digestibility improved by 4.5, 10, 16 and 20%, respectively. The improvement in IVPD of fermented flour before and after radiation is likely to be due to reduction in antinutrients especially phytate which degraded by phytase during fermentation as reported by Sokrab et al. (2012). On the other hand the reduction in IVPD may be due to disulphide formation resulting in disulphide cross-linked protein oligomers and polymers. These polymers form a coat reducing the accessibility of the protein bodies to enzymatic attack (Duodu et al., 1999).

Effect of gamma radiation and/or traditional processing on total and extractable some major and trace minerals

Table 2 shows the content and HCl extractability of some major minerals (Na, K and Mg) of high tannin sorghum cultivar (WadAhmed) as affected by radiation and/or traditional processings. For the raw flour, Na content was found to be 18.10 mg/100 g and out of this amount about 65.70% was found to be extractable; K was 380.00 mg/100 g with extractability of 51.60% and Mg was 54.30 mg/100 g and out of this amount about 47.50% was extractable. Variations in extractability between Na, K and Mg is likely to be due to the binding ability of such minerals with antinutrients which are observed to form complexes with minerals and reduced their

extractability as reported by Idris et al. (2005). Radiation of raw flour had no significant ($P \le 0.05$) effect on total Na, K and Mg but significantly (P \leq 0.05) increased the extractability of such minerals. The increment in minerals extractability after radiation could be attributed to reduction in the antinutrients. Fermentation of raw flour significantly $(P \le 0.05)$ increased both total and extractable Na, K and Mg. Fermentation leads to lowering of pH as a consequence of bacterial production of lactic and acetic acids, which is favourable for phytase activities, resulting in lowering of phytate. The reduction in phytate resulted in an improvement in minerals extractability. Cooking of raw flour decreased both total and extractable minerals. Cooking of raw fermented dough alleviates the effect of cooking and significantly ($P \le 0.05$) increased minerals extractability. Fermentation of radiated flour significantly ($P \le 0.05$) increased both total and extractable minerals. Radiation of the flour at 5, 10 or 15 KGy followed by cooking decreased minerals content but increased their extractability as compared to raw samples. Cooking of irradiated fermented dough caused a significant ($P \le 0.05$) improvement in extractable minerals. Moreover, as the level of radiation increased the degree of improvement in minerals extractability was significant (P ≤ 0.05). The results obtained showed that processing of radiated flour significantly ($P \le 0.05$) increased the extractable Na, K and Mg with increase in radiation dose but had no significant (P ≤ 0.05) effect on total minerals. Generally the increase in extractable minerals during traditional processing before and after radiation could be attributed to the reduction in the level of antinutrients as a result of such treatments (Idris et al., 2007; Mohamed Nour et al., 2010).

Table 3 shows the content and HCl extractability of some trace minerals (Zn, Cu, Mn and Co) of

Radiation dose		Minerals										
(KGy)	Treatment	Z	Zn –	(Cu	Ν	1n	Со				
		Total	extractable	Total	extractable	Total	extractable	Total	extractable			
0	Raw	2.54±1.09ª	46.66 (±0.41 ^m	0.43±0.06ª	35.83 ± 0.34^{i}	3.39±0.04b	38.12±0.02m	0.18±0.19 a	66.47±1.94 ⁿ			
	Fermented	2.62±0.11ª	64.70±0.75 ^f	$0.42{\pm}0.03^{a}$	60.00±0.62e	3.44±0.13 ^b	53.69 ± 2.02^{f}	0.20±0.11 a	83.11 ± 0.33^{f}			
	Cooked	2.54±0.21ª	41.48±0.54 ⁿ	0.45±3.17ª	29.32±1.47k	3.31±0.24b	35.66±0.36 ⁿ	0.17±1.81 a	64.79±0.75 ^p			
	Fermented/cooked	2.58±0.92ª	61.51 ± 0.02^{h}	0.46±0.05ª	49.38 ± 0.15^{f}	3.77±0.64 ^b	52.49±1.96g	0.19±1.83 a	79.90±0.81g			
5	Raw	2.53±0.01ª	48.50±0.23k	0.43±0.31ª	36.50±1.98 ⁱ	3.36±0.07b	43.80 ± 0.56^{k}	0.18±0.75 a	69.20±0.11 ^m			
	Fermented	2.58±0.80 ^a	69.60±0.76e	0.44±0.76 ^a	65.40 ± 0.34^{d}	3.38 ± 0.75^{b}	69.40±0.36°	0.19±0.75 a	85.10±0.06e			
	Cooked	2.55±0.43ª	49.40±0.341	0.45±0.23ª	33.30±0.89 ^j	3.37±0.24 ^b	39.30±0.871	0.17±0.65 a	65.80±0.08°			
	Fermented/cooked	2.55±0.76 ^a	63.50±0.23g	0.42±0.14ª	64.90 ± 0.26^{d}	3.38 ± 0.18^{b}	62.50±0.03e	0.18±0.32 a	77.30±0.01 ^h			
10	Raw	2.53±0.01ª	54.20±0.09 ^j	0.45±0.03ª	48.60 ± 0.09^{f}	3.36±0.54 ^b	48.70±1.09 ⁱ	0.18±0.04 a	72.90±0.54 ^k			
	Fermented	2.53±054ª	83.90±0.65ª	$0.48{\pm}0.98^{a}$	74.90±0.76 ^b	3.37 ± 0.09^{b}	74.70±0.27ª	0.21±0.34 a	97.90±0.65ª			
	Cooked	2.51±0.21ª	63.10±0.32g	$0.42{\pm}0.37^{a}$	44.90±0.12g	3.35±0.76 ^b	48.30±0.95 ^j	0.17±0.78 ^a	70.20±0.431			
	Fermented/cooked	2.52±0.23ª	76.60±0.37 ^d	0.46±0.28ª	72.90±0.98°	3.41±0.37 ^b	68.20±0.53d	0.19±0.39 a	85.60±0.23 ^d			
15	Raw	2.52±0.03ª	55.70 ± 0.07^{i}	0.43±0.14 ^a	46.10±0.15g	3.38±0.06 ^b	52.30±0.08g	0.17±0.15 a	76.50±0.15 ⁱ			
	Fermented	2.56±0.65ª	80.50±0.13b	0.47±0.21ª	77.40±0.03ª	3.39±0.76 ^b	72.10±0.86 ^b	0.21±0.86 a	96.90±0.60 ^b			
	Cooked	2.53±0.23ª	63.50 ± 0.34^{g}	0.47±0.65ª	43.50±0.86 ^h	3.34±0.06 ^b	49.30±0.24 ^h	0.15±0.97 ^a	74.90±0.75 ^j			
	Fermented/cooked	2.52 ^a ±0.21 ^a	78.60±0.61°	0.46 ± 0.14^{a}	75.80±0.24 ^b	3.36±0.09b	68.30±0.76 ^d	0.16±0.34 a	86.40±0.05°			

Table 3. Total (mg/100 gm) and extractable (%) trace minerals of WadAhmed cultivar as affected by gamma irradiation followed by processing

sorghum cultivar (WadAhmed) as affected by radiation and/or traditional processings. The raw flour of the cultivar contained 2.54, 0.43, 3.39 and 0.18 mg/100 g of Zn, Cu, Mn and Co, respectively and out of this amount about 46.66, 35.83, 38.12 and 66.47% were found to be extractable for the minerals, respectively. Variations in extractability between trace minerals are likely to be due to the binding ability of such minerals with antinutrients as reported by Idris et al. (2005). Radiation of raw flour had no significant ($P \le 0.05$) effect on total minerals but significantly ($P \le 0.05$) increased the HCl extractability of such minerals. Fermentation of raw flour slightly increased total minerals and significantly ($P \le 0.05$) increased the extractable ones. Cooking of raw flour had no significant ($P \le 0.05$) effect on total and extractable minerals. Cooking of fermented dough alleviates the effect of cooking and significantly ($P \le 0.05$) increased the extractability of trace minerals but did not significantly ($P \le 0.05$) affect total minerals. Radiation at all levels followed by fermentation of raw flour gave varying changes in total and extractable minerals for the cultivar but generally there is a significant ($P \le 0.05$) increase in total as well as in extractable minerals compared to raw flour. However, cooking of radiated flour had no great effect on content but significantly ($P \le 0.05$) increased extractability of trace minerals. Cooking of radiated/fermented dough caused a significant $(P \le 0.05)$ improvement in extractable minerals but did not affect total minerals. The results obtained showed that as the level of radiation increased the degree of improvement in minerals extractability was significant ($P \le 0.05$). The results showed that processing of irradiated flour significantly (P \leq (0.05) increased the extractable trace minerals with increase in radiation dose but had no significant (P

 \leq 0.05) effect on total trace minerals. The increment in extractable minerals during traditional processing before and after radiation could be attributed to the reduction in the level of antinutrients as a result of such treatments.

Effect of gamma radiation and/or traditional processing on amino acid composition and scores

Tables 4 shows the effect of radiation process on amino acids composition of raw and treated flour of WadAhmed cultivar. Radiation of raw flour of the cultivar at 5, 10 and 15 kGy decreased the level of all amino acids except leucine and phenylalanine plus tyrosine with a concomitant decrease in amino acid score except that of leucine and phenylalanine + tyrosine. Joseph et al. (2005) reported that with the exception of tyrosine the amino acids in cowpea (acidic, basic, polar and non polar amino acids) were decreased significantly with increase in gamma radiation compared to their respective controls. Hooshmand and Klopfenstein (1995) reported that lysine content decreased by 7 and 13% respectively, when maize and wheat flours were irradiated at 7.5 kGy. The observed increase in some free amino acid content due to exposure to ionizing radiation is in agreement with the findings reported by Satter et al. (1990), who documented increases in essential and nonessential amino acids of soybean when irradiated at a dose level of 10 kGy. The precise effect of ionising radiation on free amino acid content depends on various factors, such as sensitivity of the exposed system, the type of particular functional tissue and even other conditions, such as aqueous soaking after irradiation, as has been indicated by Siddhuraju et al. (2002). Wang and vonSonntage (1991) reported that sulphur-containing as well as aromatic amino acids are, in general, the most sensitive to irradiation, while

		Amino acid													
Radiation dose (KGy)	Treatment	Isoleucine		Leucine		Lysine		Methionine +Cysteine		Phenyl alanine +Tyrosine		Threonine		Valine	
		Total	AAS	Total	AAS	Total	AAS	Total	AAS	Total	AAS	Total	AAS	Total	AAS
0	Raw	3.79 ⁱ	135.36	10.77 ^h	163.20	1.71	29.48	1.71	68.40	7.70°	122.21	1.83 ^b	53.80	5.38 ^f	153.70
	Fermented	4.17 ^d	149.00	11.42 ^d	173.00	1.96ª	33.79	1.85ª	73.96	7.65 ^p	121.40	1.61 ^f	47.20	5.527°	157.90
	Cooked	4.04°	144.29	11.49°	174.10	0.96 ^k	16.55	0.96 ^j	38.40	8.32 ^h	132.20	1.04 ⁿ	30.65	5.56 ^d	158.70
	Fermented /cooked	4.42°	158.00	11.34°	171.88	1.04 ^j	17.93	0.841	33.60	8.88°	141.00	1.051	30.76	5.82°	166.30
5	Raw	2.99 ⁿ	106.70	10.72 ⁱ	162.36	1.38 ^d	23.86	1.31 ^f	52.40	7.81 ^m	124.03	1.33 ^h	39.20	4.31 ^m	123.30
	Fermented	3.22 ^m	115.10	8.72 ^m	132.10	1.58°	27.20	1.60°	64.10	8.11 ^k	128.80	1.20 ⁱ	35.32	4.34 ¹	123.90
	Cooked	3.82 ^h	136.50	10.03 ^j	152.00	0.73 ⁿ	12.61	0.70 ^k	28.10	8.86 ^d	140.60	1.68 ^d	49.30	4.87 ^h	139.20
	Fermented/cooked	3.84 ^g	137.30	7.99 ⁿ	121.00	1.05 ⁱ	18.12	0.72 ^m	28.90	7.951	126.20	1.20 ^j	35.10	3.83 ^p	109.30
10	Raw	2.53°	90.21	10.96 ^f	166.06	1.24°	21.41	1.26 ^g	50.40	8.22 ^j	130.50	1.43 ^g	42.10	4.82 ⁱ	137.80
	Fermented	4.44 ^b	158.60	12.68ª	192.10	1.18 ^f	20.30	1.49 ^d	59.50	8.28 ⁱ	131.40	1.76°	51.70	5.95 ^b	169.90
	Cooked	3.99 ^f	142.50	10.90g	165.20	0.63°	10.80	0.66°	26.40	8.96ª	142.20	1.83ª	53.90	5.31s	151.80
	Fermented /cooked	4.47ª	159.80	11.56 ^b	175.20	0.951	16.97	1.13 ⁱ	45.20	8.73°	138.60	1.19 ^k	35.00	6.14 ^a	175.40
15	Raw	2.02 ^p	72.29	10.99 ^f	166.52	1.14 ^g	19.72	0.86 ^k	34.40	7.77 ⁿ	123.30	0.93 ^p	27.20	4.10°	117.10
	Fermented	3.231	115.30	8.17 ^m	123.80	1.08 ^h	18.50	1.37°	55.00	8.41s	133.50	1.01°	29.60	4.30 ⁿ	122.80
	Cooked	3.46 ^j	123.70	9.031	136.80	0.89m	15.33	1.18 ^h	47.20	8.56 ^f	135.90	1.04 ^m	30.70	4.61 ^k	131.60
	Fermented /cooked	3.45 ^k	123.30	9.69 ^k	146.80	0.51 ^p	8.77	0.63 ^p	25.20	8.95 ^b	142.10	1.65°	48.50	4.69 ^j	134.10

Table 4. Amino acids composition (mg/100 gm protein) and score (AAS) of WadAhmed cultivar as affected by gamma radiation followed by traditional treatments

Values are means of duplicate samples. Means not sharing a common superscript(s) in column are significantly different at $p \le 0.05$.

simple amino acids could be formed by destruction of other amino acids. Bhat et al. (2008) reported that radiation is an efficient process in maintaining the nutritional potential of Mucuna pruriens seeds. Fermentation of raw flour of the culivar increased the level of isoleucine, methionine + cystein and valine (Table 4). It has been reported that fermentation of Sicklepod leaves significantly ($P \le 0.05$) increased alanine, valine, cystine, isoleucine, leucine and ammonia contents while aspartic, threonine, serine, glutamic, tyrosine, histidine, lysine and arginine contents were significantly ($P \le 0.05$) decreased (Osman et al., 2010). Radiation at 5, 10 and 15 kGy followed by fermentation significantly ($P \le 0.05$) increased the level of phenylalanine plus tyrosine of the cultivar compared to the respective control. Cooking of raw flour of the cultivar slightly increased the level of isoeucine, leucin, valine and significantly $(P \le 0.05)$ that of phenylalanine plus tyrosine. Fageer et al. (2004) observed that cooking of corn in water increased lysine, valine, leucine and phenylalanine while threonine, methionine and isoleucine were decreased. Radiation at 5, 10 and 15 kGy followed by cooking significantly ($P \le 0.05$) increased phenylalanine plus tyrosine of the cultivar. Cooking of fermented dough of radiated and non-radiated flour increased phenylalanine plus tyrosine. Osman et al. (2010) reported that cooking of the fermented Sicklepod leaves significantly ($P \le 0.05$) decreased aspartic, threonine, serine, glutamic acid, glycine, tyrosine, phenylalanine and arginine contents. The results obtained showed that the effect of radiation alone on amino acids composition was minor as both fermentation and cooking had negative effect on some amino acids. Moreover, radiation process is an ideal method in preserving material as reported for

millet flour (Mohamed et al., 2010b).

Conclusion

The observations about nutritional quality in the studied samples tend to suggest that radiation processing up to 15 kGy had little effect on their value and had slight effect on the amino acids of the flour whether raw or processed. Therefore, radiation can be applied to alleviate the severe problem of antinutrients. Moreover, when accompanied by traditional processing an improvement in protein and minerals availability was observed.

References

- AbdElhaleem, W. H., Mustafa, A. I., El Tinay, A. H. and Babiker, E. E. 2008. Effect of fermentation, maltpretreatment and cooking on antinutritional factors and in vitro protein digestibility of sorghum cultivars. Pakistan Journal of Nutrition 7: 335-341.
- Abdelseed, B.H., Abdalla, A. H., Yagoub, A.A., Mohamed, I.A. and Babiker E. E. 2011. Some nutritional attributes of selected newly developed lines of sorghum after fermentation. Journal of Agricultural Science and Technology 13: 399-409.
- Aziz, N.H., El-Far, F.M., Shahin, A.A.M. and Roushy, SM. 2007. Control of fusarium, moulds and fumonisin B1 in seeds by gamma irradiation. Food Control 18: 1337–1342.
- Bhat, R., Sridhar, K.R., Young, C., Bhagwath, A.A. and Ganesh, S. 2008. Composition and functional properties of raw and electron beam-irradiated *Mucuna pruriens* seeds. International Journal of Food Science and Technology 43: 1338–1351.
- Bhavsar, S.P., Augustine, S.K. and Kapadnis B.P. 2007. Effect of Physical and Chemical Treatments on *Campylobacter* Spiked into Food Samples. Food

Science and Technology International 13: 277-283.

- Brigide, P. and Canniatti-Brazac, A. 2006. Anti-nutrients and *in vitro* availability of iron in irradiated common beans (*Phaseolus vulgaris*). Food Chemistry 98: 85-89.
- Casarett A.P. 1996. Radiation Biology. New York: Pp.284– 312, Prentice-Hall Inc.
- Chapman, H.D. and Pratt, P.F. 1982. Method for the analysis of soil, plant and water 2nd. California University Agricultural Division, California, pp. 170-178.
- Chauhan, B.M. and Mahjan, L. 1988. Effect of natural fermentation on the extractability of minerals from pearl millet flour. Journal of Food Science 53: 1576–1577.
- Duodu, K.G., Minnaar, A. and Taylor, J.R.N. 1999. Effect of cooking and irradiation on the labile vitamins and anti-nutrient content of a traditional African sorghum porridge and Spinach relish. Food Chemistry 66: 21-27.
- El-Niely H.F.G. 2007. Effect of radiation processing on anti-nutrients, *in vitro* protein digestibility and protein efficiency ratio bioassay of legume seeds. Radiation Physics and Chemistry 76: 1050-1057.
- Eltayeb, M. M., Hassn, A. B., Mashier, S. A., Babiker, E. E. 2007. Effect of processing followed by fermentation on antinutritional factors content of pearl millet (*Pennisetum glaucum* L.) cultivars. Pakistan Journal of Nutrition 6: 463-467
- Erkan, N. and Ozden, O. 2007. The changes of fatty acid and amino acid compositions in seabream (*Sparus aurata*) during irradiation process. Radiation Physics and Chemistry 76: 1636–1641.
- Fageer, A.M., Babiker, E. E. and El Tinay, A.H. 2004. Effect of malt pretreatment and/or cooking on phytate and essential amino acids contents and in vitro protein digestibility of corn flour. Food Chemistry 88: 261– 265.
- Helinski, M.E.H., Hassan, M.M., El-Motasim, W.M., Malcolm, C.A., Knols, B.G.J. and El-Sayed B. 2008. Methodology towards a sterile insect technique field release of Anopheles arabiensis mosquitoes in Sudan: irradiation, transportation, and field cage experimentation. Malaria Journal 7: 1–10.
- Hooshmand, H. and Klopfenstein, C.F. 1995. Effect of gamma irradiation on the mycotoxin disappearance and amino acid contents on corn, wheat and soybeans with different moisture contents. Plant Foods Human Nutrition 47: 227-238.
- Idris, W. H., AbdelRahman, S. M., ELMaki, H. B., Babiker, E. E. and EL Tinay, A H. 2005. Effect of germination, fermentation and cooking on phytic acid and tannin contents and HCl-extractability of minerals of sorghum (*Sorghum biocolor*) cultivars. Journal of Food Technology 3: 410-416.
- Idris, W. H., AbdelRahman, S. M., ELMaki, H. B., Babiker, E. E. and EL Tinay, A H. 2007. Effect of malt pretreatment on HCl extractability of calcium, phosphorus and iron of sorghum (*Sorghum biocolor*) cultivars. International Journal of Food Science and Technology 42: 194–199.

- Joseph, O.A., Klasus, M., Kwaku, D. and Amanda, M. 2005. Functional properties of cowpea (*Vigna unguiculata* L. Walp) flours and pastes as affected by γ irradiation. Food Chemistry 93: 103–111.
- Lee, S., Lee M. and Song K. 2005. Effect of gammairradiation on the physicochemical properties of gluten films. Food Chemistry 92: 621–925.
- Maliwal, B.P. 1983. In vitro method to assess the nutritive value of leaf concentrate. J. Agric. Food Chemistry 31: 315–319.
- Mbarki, R., Sadok, S. and Barkallah, I. 2008. Influence of gamma irradiation on microbiological, biochemical, and textural properties of bonito (*Sarda sarda*) during chilled storage. Food Science and Technology International 14: 367-373.
- Mohamed Nour, A A, Mohamed, Ahmed, I A, Babiker, E E. and Yagoub, A A. 2010. Investigations on winter season sudanese sorghum cultivars: effect of sprouting on the nutritional value. International Journal of Food Science and Technology 45: 884-890.
- Mohamed, E.A., Ali, N. A., Ahmed, S.H., Mohamed, I.A. and Babiker E.E. 2010a. Effect of radiation process on anti-nutrients and HCl extractability of calcium, phosphorus and iron during processing and storage. Radiation Physics and Chemistry 79: 791-796.
- Mohamed, E. A., Mohamed, A. I. and Babiker, E.E. 2010b. Effects of radiation process on total protein and amino acids composition of raw and processed pearl millet flour during storage. International Journal of Food Science and Technology 45: 906–912.
- Mohiedeen, I. E., Abdullahi, H. E., Abd Elmoneim, O. E., Babiker, E. E. and Mallasy, L. O. 2010. Effect of fermentation and cooking on protein quality of maize (*Zea Mays* Linnaus) cultivars. International Journal of Food Science and Technology 45, 1284-1290.
- Monjula, S. and John, E. 1991. Biochemical changes and *in vitro* protein digestibility of the endosperm of germinating Dolichos lablab. Journal of Science of Food and Agriculture 55: 229–238.
- Moore, S. and STAIN W.H. 1963. Chromatographic amino acids determination by the use of automatic recording equipment methods. Enzymolgy 63: 819–831.
- Osman, A. M., Hassan, A. B., Osman, G. A. M., Nagat, M., Mohamed, A. H., Eiman, E. D. and Babiker E. E. 2012. Effects of gamma irradiation and/or cooking on nutritional quality of faba bean (*Vicia faba* L.) cultivars seeds. Jounal of Food Science and Technology DOI10.1007/s13197-012-0662-7.
- Osman M. A. 2004. Changes in sorghum enzyme inhibitors, phytic acid, tannins and *in vitro* protein digestibility occurring during Khamir (local bread) fermentation. Food Chemistry 88: 129-134.
- Osman, N. M., Mohamed, A. I. and Babiker, E.E. 2010. Fermentation and cooking of Sicklepod (*Cassia* obtusifolia) leaves: Changes in chemical and amino acid composition, antinutrients and protein fractions and digestibility. International Journal of Food Science and Technology 45: 124-132.
- Price, M.L. and Butler, L.G. 1978. Rapid visual estimation and spectrophotometric determination of tannin

content of sorghum grain. Journal of Agriculture and Food Chemistry 25: 1268–1273.

- Satter, A. and Akhtar, M.A. 1990. Irradiation and germination effects on phytate, protein and amino acids of soybean. Plant Foods Human Nutrition 40: 185–195.
- Sendra, E., Capellas, M., Guamis, B., Felipe, X., Mor-Mur, M. and Pla, R. 1996. Revisión: Irradiación de alimentosaspectos generales/Review: Food irradiation-general aspects. Food Science and Technology International 2: 1-11.
- Siddhuraju, P., Makkar, H.P.S. and Becker, K. 2002. The effect of ionizing radiation on antinutritional factors and the nutritional value of plant materials with reference to human and animal food. Food Chemistry 78: 187–205.
- Snedecor, G.W. and Cochran, W.G. 1987. Statistical Methods, 17th edn. Pp. 221–222. Ames, IA: The Iowa State University Press.
- Sokrab, A. M., Isam, A. M. and Babiker, E. E. 2012. Effect of malting and fermentation on antinutrients, and total and extractable minerals of high and low phytate corn genotypes. International Journal of Food Science and Technology 47: 1037-1043.
- Wang, D. and vonSonntage, C. 1991. Radiation industrial oxidation of phenylalanine. In: Proceedings of the Workshop on Recent Advances on Detection of Irradiated Foods, BRC Information, Chem. Anal. Comm. of the Eur. Comm., Brussels. Pp. 212–217.
- Wheeler, E.L. and Ferrel, R.E. 1971. A method for phytic acid determination in wheat and wheat fractions. Cereal chemistry 48: 312–320.
- WHO 1999. High-dose Irradiation: Wholesomeness of Food Irradiated with Doses above 10 kGy. Geneva: WHO (WHOTechnicalReport-Series no, 890).